

MONOCLONAL ANTIBODIES AGAINST HUMAN COLON CARCINOMA- ASSOCIATED ANTIGENS AND USES THEREFOR

This application is a continuation-in-part of U.S. application Ser. No. 08/159,836 filed Nov. 30, 1993, which is a continuation-in-part of U.S. application Ser. No. 08/117,430, filed Sep. 7, 1993, now abandoned, which is a continuation-in-part of U.S. application Ser. No. 07/670,816, filed Mar. 18, 1991, now abandoned, which is a continuation-in-part of U.S. application Ser. No. 07/176,337, filed Mar. 31, 1988, now abandoned.

BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention, in the field of immunology and medicine, relates to new hybridoma lines and the monoclonal antibodies (mAbs) they secrete which are specific for clinically defined colon carcinoma-associated antigens. The mAbs are useful in vivo for immunodetection and immunotherapy of colon carcinoma as well as for the detection and purification of colon carcinoma-associated antigens.

2. Description of the Background Art

During the process of oncogenesis, a number of cell-surface molecules or markers appear on cells. Such tumor-related markers include oncofetoproteins, neoglycoproteins, sphingolipids, and modifications of existing surface proteins. Such new or altered structures are often shed from the tumor cell surface and appear in the serum or in other biological fluids. The detection of any of these substances or "tumor markers" serves as the basis for diagnosing or monitoring the progress of neoplastic disease.

Early animal studies demonstrated that, among these tumor markers, a subset of tumor membrane protein or glycoprotein antigens were immunogenic. Upon appropriate reintroduction into the tumor-bearing host, typically after surgical removal of the primary tumor, such antigens could effectively block the establishment of new tumor growth.

An attempt to use similarly derived tumor-associated antigens in humans was made by Hollinshead and Stewart using a relatively purified membrane preparation in patients with lung cancer (Stewart, T. H. M. et al., *Ann. N.Y. Acad. Sci.* 277:436 (1976)). These studies were later expanded to include patients with melanoma and colon carcinoma, wherein different pooled allogeneic tumor preparations were administered in combination with complete Freund's adjuvant (Hollinshead, A. C. et al., *Cancer* 4:9:1387 (1982); Hollinshead, A. C. et al., *Cancer* 56:480 (1985)).

The use of Freund's adjuvant was based on observations that normal tissue antigens with this adjuvant produced severe autoimmune responses in animal recipients, whereas in the absence of this adjuvant, no adverse reactions were seen. The adjuvant was thought to promote antigen processing by host macrophages as well as prolong the stimulatory action of the antigen at the site of its deposition (see, for example, Roitt, L., *Essential Immunology*, 6th Ed., Blackwell Scientific Publications, Oxford (1988)).

The above observations served as the basis for early clinical trials using specific human tumor membrane proteins and glycoproteins as tumor "vaccines." Various of the tumor-associated antigens which had been isolated and characterized could prolong survival and, in some cases, produce regression of metastatic disease.

With the advent of monoclonal antibody (mAb) technology, it has become possible to obtain pure antibody

populations which permit better purification and characterization of the various tumor markers and tumor-associated antigens that are useful for immunodiagnosis or immunotherapy. Many mAbs have been described that have varying degrees of selectivity for tumor antigens (versus normal cell surface markers); some of these tumor antigens are broadly represented across several or many tumor types, whereas others appear to be truly tumor-specific. A number of these mAb-tumor antigen systems are described below.

Herlyn et al., *Proc. Natl. Acad. Sci. USA* 76:1438 (1979), discloses two mAbs obtained by immunizing mice with human colorectal carcinoma (CRC) cells. The mAbs have selective reactivity with human CRC cells. One mAb, 1083-17 (the forerunner of 17-1A), is now known to react with a 41 kDa glycoprotein (see below).

Herlyn et al., *J. Clin. Immunol.* 2:135 (1982), described the detection of a circulating colorectal carcinoma (CRC)-associated antigen by a mAb developed against a membrane antigen of the SW116 cell line. MABs 19-9 and 52a, which recognize a monosialoganglioside antigen (Magnani, J. L. et al., *Science* 212:55 (1981)), reacted with cells of 8 of 12 CRC lines as well as with the cells of one gastric carcinoma and one pancreatic carcinoma. MAB C₄14 reacted with four of six CRC cultures and with gastric tumor cells. The binding of mAbs 19-9 and 52a to tumor cells was inhibited by a CRC patient's serum. However, CRC sera inhibited binding less frequently than did sera from patients with pancreatic or gastric tumors.

Girardet et al., *J. Immunol.* 136:1497-1503 (1986), disclosed mAbs against human colon carcinomas. The L-D1 mAb reacted with a 41 kDa glycoprotein, believed to be the same antigen as that defined by mAb 1083-17-1A (Herlyn et al., 1979, supra). The L-C5 mAb precipitated proteins having molecular weights of 43, 45, 47 and 53 kDa from LoVo colon carcinoma cells. L-D1 did react with cervical carcinoma lines, while L-C5 reacted with breast carcinoma lines. Their binding to pancreatic carcinomas was not examined.

Greiner et al., *Science* 235:895-898 (1987), discloses mAb 06.2 which reacts with a 90 kDa glycoprotein allegedly found in 75-80% of breast carcinomas and more than 90% of colon carcinomas.

Sakamoto et al., U.S. Pat. No. 4,579,827 (Apr. 1, 1986), discloses a number of mAbs said to be useful for diagnosing or treating human colon cancer by a number of different approaches. None of these mAbs is shown to react with a human colon carcinoma-associated antigen that is a protein of either 61 or 72 kDa molecular weight, distinguishing these antibodies from the antibodies of the present invention (described below). Furthermore, none of the Sakamoto mAbs have the degree of colon tumor specificity of the mAbs disclosed in the present application.

Delaloye et al., *J. Clin. Invest.* 77:301 (1986), discloses the use of a mAb specific for carcinoembryonic antigen (CEA) to detect colorectal carcinoma in vivo using ¹²⁵I-labelled fragments and emission computerized tomography. The mAb described bears no relation to the mAbs of the present invention.

Douillard et al., *Hybridoma* 5, Suppl. 1:S139 (1986), describes mAb 17-1A and its cytotoxic properties to gastrointestinal adenocarcinomas in vitro. 17-1A was used with some degree of success in immunotherapy trials. This mAb is said to recognize a 38-41 kDa protein and has a broad range of reactivity and lack of colon tumor specificity, clearly distinguishing it from the antibodies of the present invention.

Scannon et al., U.S. Pat. No. 4,590,071 (May 20, 1986), discloses mAbs specific for melanoma antigens conjugated

to toxic proteins such as ricin A chain and the use of these compositions to treat melanoma. There is no disclosure directed to colon tumor antigens or antibodies and their uses.

The relatively pure antigen preparation containing the immunogenic colon carcinoma membrane antigens to which the mAbs of the present invention are directed was originally described by Hollinshead et al., *Science* 177:887-889 (1972).

The clinical evaluation of the above antigen preparation, including a description of its immunogenicity and potential for enhancing patient survival through stimulation of specific active immunity, was described by Hollinshead et al., 1985, *supra*.

The work of the present inventors leading to the present invention is briefly described in an abstract by Tsang et al., "Monoclonal Antibodies to Human Colon Carcinoma Associated Antigens," Intl. Symp. Biotech. in Clin. Med., Rome, Italy, Apr. 13-15, 1987. This reference was made available to the public less than one year before the filing of the ultimate parent application (U.S. Ser. No. 07/176,337) for the present application.

SUMMARY OF THE INVENTION

The present inventors have produced murine mAbs and mouse-human chimeric antibodies specific for colon carcinoma-associated antigens which were known to be immunogenic in humans. These antigens, isolated in the inventors' laboratory, are unique among the previously described colon cancer antigens in that (1) the epitopes recognized by the mAbs are of the protein and not the carbohydrate component of tumor-associated glycoproteins; (2) the antigens are not expressed in normal tissues; (3) the antigens are tumor-specific, being present in malignancies of colon, breast, and ovarian cancer; (4) the antigens are immunogenic in humans, having the capability of enhancing host anti-tumor immunity by both cellular as well as humoral responses, thus improving survival in cancer patients; and (5) the immunogenicity in humans is specific, in that only colon, breast and ovarian cancer patients, but not patients with other forms of cancer, show evidence of specific *in vivo* or *in vitro* immunological reactivity to the antigens.

The mAbs and chimeric antibodies of the present invention are useful for diagnosis or therapy of colon, breast, and ovarian carcinoma, for example by imaging metastatic tumors, by delivering cytotoxic agents to the tumors, and by activating host effector mechanisms such as antibody-dependent cellular cytotoxicity (ADCC) or complement dependent cytotoxicity (CDC) to directly kill tumor cells.

The present invention is thus directed to a monoclonal antibody specific for a human colon carcinoma-associated protein antigen wherein the antigen is specifically immunogenic in humans, and the antigen is not detectable on normal human tissues or on human carcinoma cells other than colon, breast and ovarian carcinoma cells. Also included are antigen-binding fragments or derivatives of the antibody.

The present invention is also directed to a chimeric antibody specific for a human colon carcinoma-associated protein antigen wherein the antigen is not detectable on normal human tissues or on human carcinoma cells other than colon carcinoma cells. Mouse hybridoma PCA 31.1 has been deposited at ATCC and assigned ATCC HB-12314. Mouse hybridoma PCA 33.28 has been deposited at ATCC and assigned ATCC HB-12315. Cells transfected with chimeric 31.1 have been deposited at ATCC and assigned ATCC CRL-12316. The above deposits were made at

American Type Culture Collection, 12301 Parklawn Drive, Rockville, Md. 20862 USA on Mar. 13, 1997.

In one embodiment, the antibody is specific for a CCAA which is a protein having a molecular weight of about 61 kilodaltons. In another embodiment, the antibody is specific for a CCAA which is a protein having a molecular weight of about 72 kilodaltons. In a preferred embodiment, the antibody is the mouse monoclonal antibody 33.28 or 31.1 or an antibody which binds specifically to the same colon carcinoma-associated epitope as that bound by 33.28 or 31.1. In another preferred embodiment, the antibody is a mouse/human chimeric antibody Chi #1 that binds specifically to the same colon carcinoma-associated epitope as that bound by 31.1.

The present invention is also directed to the above antibody immobilized on a solid phase.

The present invention includes the above antibody detectably labelled, for example, with a radiolabel.

In additional embodiments, the above antibody is conjugated to a cytotoxic radionuclide, a cytotoxic drug, or a cytotoxic protein.

In yet another embodiment, the present invention is directed to monoclonal antibodies against the above antibodies, i.e., second generation monoclonal antibodies.

In a further embodiment, the present invention is directed to third generation monoclonal antibodies, i.e., monoclonal antibodies directed against the above second generation monoclonal antibodies.

The present invention is also directed to the above-discussed colon carcinoma-associated antigens which are unique in that (1) the epitopes recognized by the mAbs are of the protein and not the carbohydrate component of tumor-associated glycoproteins; (2) the antigens are not expressed in normal tissues; (3) the antigens are tumor-specific, being present in the malignancies of colon, breast and ovarian cancer; (4) the antigens are immunogenic in humans, having the capability of enhancing host anti-tumor immunity, thus improving survival in cancer patients; and (5) the immunogenicity in humans is specific, in that only colon, breast and ovarian cancer patients, but not patients with other forms of cancer, show evidence of specific *in vivo* or *in vitro* immunological reactivity to the antigens.

To date, all other purified antigens that have been used have failed to elicit both a cellular and a humoral response.

The present invention also provides a pharmaceutical composition useful for the immunotherapy of colon, breast and ovarian cancer comprising an antibody, fragment or derivative, as above, conjugated to a cytotoxic radionuclide, a cytotoxic drug, or a cytotoxic protein, in a suitable excipient.

The present invention includes an immunoassay method for detecting in a sample a colon carcinoma-associated antigen capable of binding to the 33.28 or 31.1 murine monoclonal antibody or Chi #1, comprising:

- (a) contacting the sample with an antibody described above; and
- (b) detecting the antigen by detecting the binding of the antibody.

In another embodiment, the invention provides an imaging method for detecting a colon carcinoma-associated antigen in a subject, comprising:

- (a) contacting the detectably labelled antibody as described above with the subject; and
- (b) detecting the antigen.

The present invention also includes a method of killing cells carrying a colon carcinoma-associated antigen, comprising:

- groupings of molecules, for example, amino acids or sugar side chains, and have specific three-dimensional structural characteristics as well as specific charge characteristics. The epitopes of interest for the present invention are epitopes comprising amino acids.

An "antigen" is a molecule or a portion of a molecule capable of being bound by an antibody which is additionally capable of inducing an animal to produce an antibody capable of binding to an epitope of that antigen. An antigen may have one or more than one epitope. The specific reaction referred to above is meant to indicate that the antigen will react, in a highly selective manner, with its corresponding antibody and not with the multitude of other antibodies which may be evoked by other antigens.

The term "antibody" is meant to include both intact immunoglobulin molecules as well as fragments and derivatives thereof, such as, for example, Fab, Fab', F(ab')₂ and Fv, which are capable of binding antigen. These fragments lack the Fc fragment of intact antibody, clear more rapidly from the circulation, and may have less non-specific tissue binding than an intact antibody (Wahl et al., *J. Nucl. Med.* 24:316-325 (1983)). These fragments are produced from intact antibodies using methods well known in the art, for example by proteolytic cleavage with enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')₂ fragments).

A "derivative" of an antibody contains additional chemical moieties not normally a part of the protein. Covalent modifications of the protein are included within the scope of this invention. Such modifications may be introduced into the molecule by reacting targeted amino acid residues of the antibody with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues. For example, derivatization with bifunctional agents, well-known in the art, is useful for cross-linking the antibody or fragment to a water-insoluble support matrix or to other macromolecular carriers.

- By "vaccine" is meant an agent used to stimulate the immune system of a living organism so that immunological protection against future harm caused by an infectious agent is provided. Administration of a vaccine contemplated by the present invention to the patient may be by any known or standard techniques. These include oral ingestion, intestinal intubation, or broncho-nasal spraying. Other methods of administration, such as intravenous injection, that allow the carrier microbe to reach the human or animal's bloodstream may be acceptable when the carrier microbe is unable to reproduce.

- The antibodies of the present invention are novel in that they are the first known mAbs and chimeric antibody specific for CCAA wherein the tumor antigens are known to be immunogenic in humans. That is, the antigens recognized by the antibodies of the present invention induce an immune response in patients with colon, breast, and ovarian cancer, but not in other individuals, such as patients with other types of cancer. The immunogenicity of these antigens is expressed chiefly as cell-mediated immunity, measurable either by assay of delayed cutaneous hypersensitivity in vivo ("skin tests"), or by various in vitro assays of specific lymphocyte reactivity, such as lymphocyte proliferation or lymphocyte migration inhibition assays. For general principles of immunogenicity and description of various assays of specific immunological reactivity, see: Roitt, L. *Essential Immunology*, 6th Ed., Blackwell Scientific Publications, Oxford (1988); Roitt, I. et al., *Immunology*, C. V. Mosby Co., St. Louis, Mo. (1985); Klein, J., *Immunology*, Black-

individual polypeptide chains, as taught, for example by Sears et al., *Proc. Natl. Acad. Sci. USA* 72:353-357 (1975). With this approach, hosts expressing chimeric H chains (or their derivatives) are separately cultured from hosts expressing chimeric L chains (or their derivatives) and the immunoglobulin chains are separately recovered and then associated. Alternatively, the hosts can be co-cultured and the chains allowed to associate spontaneously in the culture medium, followed by recovery of the assembled immunoglobulin, fragment or derivative.

Murine hybridomas which produce mAb specific for CCAA, such as the 33.28 and 31.1 mAbs of the present invention, are formed by the fusion of a mouse fusion partner cell, such as SP2/0, and spleen cells from mice immunized against the CCAA.

Mice may be immunized with crude or semi-purified preparations containing the antigens of interest, such as, for example, the Hollinshead "vaccine," which is a partially purified membrane preparation of colon-carcinoma cells (Hollinshead et al., *supra*). To immunize the mice, a variety of different conventional protocols may be followed. For example, mice may receive primary and boosting immunizations of antigenic preparations.

The cell fusions are accomplished by standard procedures well known to those skilled in the field of immunology (Kohler and Milstein, *Nature* 256:495-497 (1975) and U.S. Pat. No. 4,376,110; Hartlow, E. et al., supra; Campbell, A., "Monoclonal Antibody Technology." In: *Laboratory Techniques in Biochemistry and Molecular Biology*, Volume 13 (Burdon, R. et al., eds.), Elsevier, Amsterdam (1984); Kennett et al., *Monoclonal Antibodies* (Kennett et al., eds. pp. 365-367, Plenum Press, N.Y., 1980); de St. Groth, S. F., et al., *J. Immunol. Meth.* 35:1-21 (1980); Galfre, G. et al., *Methods Enzymol.* 73:3-46 (1981); Goding, J. W. 1987, *Monoclonal Antibodies: Principles and Practice*, 2nd ed. Academic Press, London, 1987).

Fusion partner cell lines and methods for fusing and selecting hybridomas and screening for mAbs are well known in the art (Hardlow, E. et al., supra; Kawamoto, T. et al., *Meth. Enzymol.* 121:266-277 (1986); Kearney, J. F. et al., *J. Immunol.* 123:1548-1550 (1979); Kilmartin, J. V. et al., *J. Cell Biol.* 93:576-582 (1982); Kohler, G. et al., *Eur. J. Immunol.* 6:292-295 (1976); Lane, D. P. et al., *J. Immunol. Meth.* 47:303-307 (1981); Mueller, U. W. et al., *J. Immunol. Meth.* 87:193-196 (1986); Pontecorvo, G., *Somatic Cell Genet.* 1:397-400 (1975); Sharo, J., et al., *Proc. Natl. Acad. Sci. USA* 7:6:1420-1424 (1979); Shulman, M. et al., *Nature* 276:269-270 (1978); Springer, T. A. (ed), *Hybridoma Technology in the Biosciences and Medicine*. Plenum Press, New York, 1985; and Taggart, R. T. et al., *Science* 219:1228-1230 (1982)).

The mAbs of the present invention may be produced in large quantities by injecting hybridoma cells secreting the antibody into the peritoneal cavity of mice and, after appropriate time, harvesting the ascites fluid which contains a high titer of the mAb, and isolating the mAb therefrom. Alternatively, the mAbs may be produced by culturing hybridoma cells in vitro and isolating the secreted mAb from the cell culture medium.

Human genes which encode the C regions of the chimeric antibodies of the present invention are derived from cells which express, and preferably, produce, human immunoglobulins. The human C_H region can be derived from any of the known classes or isotypes of human H chains, including gamma, μ , α , δ or ϵ . Since the H chain isotype is responsible for the various effector functions of an antibody, the choice

Antibodies, fragments or derivatives having chimeric H chains and L chains of the same or different V region binding specificity can be prepared by appropriate association of the

of C_H region will be guided by the desired effector functions, such as complement fixation, or activity in antibody-dependent cellular cytotoxicity (ADCC). Preferably, the C_H region is derived from gamma 1 (IgG1), gamma 3 (IgG3), gamma 4 (IgG4), and μ (IgM).

The human C_L region can be derived from either human L chain isotype, kappa or lambda.

Genes encoding human immunoglobulin C regions are obtained from human cells by standard cloning techniques (Sambrook, J. et al., *Molecular Cloning: A Laboratory Manual*, 2nd Edition, Cold Spring Harbor Press, Cold Spring Harbor, N.Y. (1989)). Human C region genes are readily available from known clones containing genes representing the two classes of L chains and the five classes of H chains. Chimeric antibody fragments, such as $F(ab')_2$ and Fab, can be prepared by designing a chimeric H chain gene which is appropriately truncated. For example, a chimeric gene encoding the H chain portion of an $F(ab')_2$ fragment would include DNA sequences encoding the CH_1 domain and hinge region of the H chain, followed by a translational stop codon to yield the truncated molecule.

Generally, the chimeric antibodies of the present invention are produced by cloning DNA segments encoding the H and L chain antigen-binding regions of a CCAA-specific antibody, preferably non-human, most preferably 33.28 or 31.1, and joining these DNA segments to DNA segments encoding human C_H and C_L regions to produce chimeric immunoglobulin-encoding genes.

Thus, in a preferred embodiment, a fused gene is created which comprises a first DNA segment that encodes at least the antigen-binding region of non-human origin, such as a functionally rearranged V region with joining (J) segment, linked to a second DNA segment encoding at least a part of a human C region. This fusion can be accomplished by the polymerase chain reaction, as reported by Fernando et al., *Miami Symp. Short Reports* 3: 88, 1993.

The DNA encoding the antibody-binding region may be genomic DNA or cDNA. A convenient alternative to the use of chromosomal gene fragments as the source of DNA encoding the murine V region antigen-binding segment is the use of cDNA for the construction of chimeric immunoglobulin genes, as reported by Liu et al., *Proc. Natl. Acad. Sci., USA* 84:3439 (1987) and *J. Immunology* 139:3521 (1987), which references are hereby incorporated by reference. The use of cDNA requires that gene expression elements appropriate for the host cell be combined with the gene in order to achieve synthesis of the desired protein. The use of cDNA sequences is advantageous over genomic sequences (which contain introns), in that cDNA sequences can be expressed in bacteria or other hosts which lack appropriate RNA-splicing systems.

Therefore, in an embodiment utilizing cDNA encoding the antibody V region, the method of producing the chimeric antibody involves several steps, outlined below:

1. Isolation of messenger RNA (mRNA) from the cell line producing the monoclonal antibody, cloning and cDNA production therefrom;
2. Preparation of a full length cDNA library from purified mRNA from which the appropriate V region gene segments of the L and H chain genes can be: (i) identified with appropriate probes, (ii) sequenced, and (iii) made compatible with a C gene segment;
3. Preparation of C region gene segments by cDNA preparation and cloning;
4. Construction of complete H or L chain coding sequences by linkage of the cloned specific V region

gene segments to cloned human C region gene, as described above;

5. Expression and production of chimeric L and H chains in selected hosts, including prokaryotic and eukaryotic cells.

One common feature of all immunoglobulin H and L chain genes and their encoded mRNAs is the J region. H and L chain J regions have different sequences, but a high degree of sequence homology exists (greater than 80%) among each group, especially near the C region. This homology is exploited in this method and consensus sequences of H and L chain J regions may be used to design oligonucleotides for use as primers for introducing useful restriction sites into the J region for subsequent linkage of V region segments to human C region segments.

C region cDNA vectors prepared from human cells can be modified by site-directed mutagenesis to place a restriction site at the analogous position in the human sequence. For example, one can clone the complete human kappa chain C (C_k) region and the complete human gamma-1 C region ($C_{\gamma-1}$). In this case, the alternative method based upon genomic C region clones as the source for C region vectors would not allow these genes to be expressed in bacterial systems where enzymes needed to remove intervening sequences are absent. Cloned V region segments are excised and ligated to L or H chain C region vectors. Alternatively, the human $C_{\gamma-1}$ region can be modified by introducing a termination codon thereby generating a gene sequence which encodes the H chain portion of an Fab molecule. The coding sequences with linked V and C regions are then transferred into appropriate expression vehicles for expression in appropriate hosts, prokaryotic or eukaryotic.

Two coding DNA sequences are said to be "operably linked" if the linkage results in a continuously translatable sequence without alteration or interruption of the triplet reading frame. A DNA coding sequence is operably linked to a gene expression element if the linkage results in the proper function of that gene expression element to result in expression of the coding sequence.

Expression vehicles include plasmids or other vectors. Preferred among these are vehicles carrying a functionally complete human C_H or C_L chain sequence having appropriate restriction sites engineered so that any V_H or V_L chain sequence with appropriate cohesive ends can be easily inserted therein. Human C_H or C_L chain sequence-containing vehicles thus serve as intermediates for the expression of any desired complete H or L chain in any appropriate host.

A chimeric mouse-human antibody will typically be synthesized from genes driven by the chromosomal gene promoters native to the mouse H and L chain V regions used in the constructs; splicing usually occurs between the splice donor site in the mouse J region and the splice acceptor site preceding the human C region and also at the splice regions that occur within the human C_H region; polyadenylation and transcription termination occur at native chromosomal sites downstream of the human coding regions.

Gene expression elements useful for the expression of cDNA genes include: (a) viral transcription promoters and their enhancer elements, such as the SV40 early promoter (Okayama, H. et al., *Mol. Cell. Biol.* 3:280 (1983)), Rous sarcoma virus LTR (Gorman, C. et al., *Proc. Natl. Acad. Sci., USA* 79:6777 (1982)), and Moloney murine leukemia virus LTR (Grosschedl, R. et al., *Cell* 41:885 (1985)); (b) splice regions and polyadenylation sites such as those derived from the SV40 late region (Okayama et al., supra); and (c) polyadenylation sites such as in SV40 (Okayama et al., supra).

Immunoglobulin cDNA genes may be expressed as described by Liu et al., Supra, and Weidle et al., *Gene* 51:21 (1987), using as expression elements the SV40 early promoter and its enhancer, the mouse immunoglobulin H chain promoter enhancers, SV40 late region mRNA splicing, rabbit β -globin intervening sequence, immunoglobulin and rabbit β -globin polyadenylation sites, and SV40 polyadenylation elements. For immunoglobulin genes comprised of part cDNA, part genomic DNA (Whittle et al., *Protein Engineering* 1:499 (1987)), the transcriptional promoter may be human cytomegalovirus (CMV), the promoter enhancers derived from CMV and mouse/human immunoglobulin, and mRNA splicing and polyadenylation regions derived from the native chromosomal immunoglobulin sequences.

In one embodiment, for expression of cDNA genes in rodent cells, the transcriptional promoter is a viral LTR sequence, the transcriptional promoter enhancers are either or both the mouse immunoglobulin heavy chain enhancer and the viral LTR enhancer, the splice region contains an intron of greater than 31 bp, and the polyadenylation and transcription termination regions are derived from the native chromosomal sequence corresponding to the immunoglobulin chain being synthesized. In other embodiments, cDNA sequences encoding other proteins are combined with the above-recited expression elements to achieve expression of the proteins in mammalian cells.

Each fused gene is assembled in, or inserted into, an expression vector. Recipient cells capable of expressing the chimeric immunoglobulin chain gene product are then transfected singly with a chimeric H or chimeric L chain-encoding gene, or are co-transfected with a chimeric H and a chimeric L chain gene. The transfected recipient cells are cultured under conditions that permit expression of the incorporated genes and the expressed immunoglobulin chains or intact antibodies or fragments are recovered from the culture. In one embodiment, the fused genes encoding the chimeric H and L chains, or portions thereof, are assembled in separate expression vectors that are then used to co-transfect a recipient cell.

Each vector may contain two selectable genes, a first selectable gene designed for selection in a bacterial system and a second selectable gene designed for selection in a eukaryotic system, wherein each vector has a different pair of genes. This strategy results in vectors which first direct the production, and permit amplification, of the fused genes in a bacterial system. Subsequently, the genes so produced and amplified in a bacterial host are subsequently used to co-transfect a eukaryotic cell, and allow selection of a co-transfected cell carrying the desired transfected genes.

Examples of selectable genes of use in a bacterial system are the gene that confers resistance to ampicillin and the gene that confers resistance to chloramphenicol. Preferred selectable genes for use in eukaryotic transfectants include the xanthine guanine phosphoribosyl transferase gene (designated gpt) and the phosphotransferase gene from Tn5 (designated neo). Selection of cells expressing gpt is based on the fact that the enzyme encoded by this gene utilizes xanthine as a substrate for purine nucleotide synthesis, whereas the analogous endogenous enzyme cannot. In a medium containing (1) mycophenolic acid, which blocks the conversion of inosine monophosphate to xanthine monophosphate, and (2) xanthine, only cells expressing the E gene can survive. The product of the neo blocks the inhibition of protein synthesis by the antibiotic G418 and other antibiotics of the neomycin class.

The two selection procedures can be used simultaneously or sequentially to select for the expression of immunoglobulin

chain genes introduced on two different DNA vectors into a eukaryotic cell. It is not necessary to include different selectable markers for eukaryotic cells; an H and an L chain vector, each containing the same selectable marker can be co-transfected. After selection of the appropriately resistant cells, the majority of the clones will contain integrated copies of both H and L chain vectors.

Alternatively, the fused genes encoding the chimeric H and L chains can be assembled on the same expression vector.

For transfection of the expression vectors and production of the chimeric antibody, the preferred recipient cell line is a myeloma cell. Myeloma cells can synthesize, assemble and secrete immunoglobulins encoded by transfected immunoglobulin genes and possess the mechanism for glycosylation of the immunoglobulin. A particularly preferred recipient cell is the Ig-non-producing myeloma cell SP2/0 (ATCC #CRL 8287). SP2/0 cells produce only immunoglobulin encoded by the transfected genes. Myeloma cells can be grown in culture or in the peritoneal cavity of a mouse, where secreted immunoglobulin can be obtained from ascites fluid. Other suitable recipient cells include lymphoid cells such as B lymphocytes of human or non-human origin, hybridoma cells of human or non-human origin, or interspecies heterohybridoma cells.

The expression vector carrying a chimeric antibody construct of the present invention may be introduced into an appropriate host cell by any of a variety of suitable means, including such biochemical means as transformation, transfection, conjugation, protoplast fusion, calcium phosphate-precipitation, and application with polycations such as diethylaminoethyl (DEAE) dextran, and such mechanical means as electroporation, direct microinjection, and microprojectile bombardment (Johnston et al., *Science* 240:1538 (1988)). A preferred way of introducing DNA into lymphoid cells is by electroporation (Potter et al., *Proc. Natl. Acad. Sci. USA* 81:7161 (1984); Yoshikawa, K. et al., *Jpn. J. Cancer Res.* 77:1122-1133). In this procedure, recipient cells are subjected to an electric pulse in the presence of the DNA to be incorporated. Typically, after transfection, cells are allowed to recover in complete medium for about 24 hours, and are then seeded in 96-well culture plates in the presence of the selective medium. G418 selection is performed using about 0.4 to 0.8 mg/ml G418. Mycophenolic acid selection utilizes about 6 μ g/ml plus about 0.25 mg/ml xanthine. The electroporation technique is expected to yield transfection frequencies of about 10^{-5} to about 10^{-4} for Sp2/0 cells. In the protoplast fusion method, lysozyme is used to strip cell walls from catarrhal harboring the recombinant plasmid containing the chimeric antibody gene. The resulting spheroplasts are fused with myeloma cells with polyethylene glycol.

The chimeric immunoglobulin genes of the present invention can also be expressed in nonlymphoid mammalian cells or in other eukaryotic cells, such as yeast, or in prokaryotic cells, in particular bacteria.

Yeast provides substantial advantages for the production of immunoglobulin H and L chains. Yeasts carry out post-translational peptide modifications including glycosylation. A number of recombinant DNA strategies now exist which utilize strong promoter sequences and high copy number plasmids which can be used for production of the desired proteins in yeast. Yeast recognizes leader sequences of cloned mammalian gene products and secretes peptides bearing leader sequences (i.e., pre-peptides) (Hitzman, et al., 11th International Conference on Yeast Genetics and Molecular Biology, Montpellier, France, Sep. 13-17, 1982).

Yeast gene expression systems can be routinely evaluated for the levels of production, secretion and the stability of chimeric H and L chain proteins and assembled chimeric antibodies. Any of a series of yeast gene expression systems incorporating promoter and termination elements from the actively expressed genes coding for glycolytic enzymes produced in large quantities when yeasts are grown in media rich in glucose can be utilized. Known glycolytic genes can also provide very efficient transcription control signals. For example, the promoter and terminator signals of the phosphoglycerate kinase (PGK) gene can be utilized. A number of approaches may be taken for evaluating optimal expression plasmids for the expression of cloned immunoglobulin cDNAs in yeast (see Glover, D. M., ed., *DNA Cloning*, Vol. II, pp. 45-66, IRL Press, 1985).

Bacterial strains may also be utilized as hosts for the production of antibody molecules or antibody fragments described by this invention. *E. coli* K12 strains such as *E. coli* W3110 (ATCC 27325), and other enterobacteria such as *Salmonella typhimurium* or *Serratia marcescens*, and various *Pseudomonas* species may be used.

Plasmid vectors containing replicon and control sequences which are derived from species compatible with a host cell are used in connection with these bacterial hosts. The vector carries a replication site, as well as specific genes which are capable of providing phenotypic selection in transformed cells. A number of approaches may be taken for evaluating the expression plasmids for the production of chimeric antibodies or antibody chains encoded by the cloned immunoglobulin cDNAs in bacteria (see Glover, D. M., ed., *DNA Cloning*, Vol. I, IRL Press, 1985).

Other preferred hosts are mammalian cells, grown in vitro or in vivo. Mammalian cells provide post-translational modifications to immunoglobulin protein molecules including leader peptide removal, folding and assembly of H and L chains, glycosylation of the antibody molecules, and secretion of functional antibody protein.

Mammalian cells which may be useful as hosts for the production of antibody proteins, in addition to the cells of lymphoid origin described above, include cells of fibroblast origin, such as Vero (ATCC CRL 81) or CHO-K1 (ATCC CRL 61).

Many vector systems are available for the expression of cloned H and L chain genes in mammalian cells (see Glover, D. M., ed., *DNA Cloning*, Vol. II, pp. 143-238, IRL Press, 1985). Different approaches can be followed to obtain complete H₂L₂ antibodies. As discussed above, it is possible to co-express H and L chains in the same cells to achieve intracellular association and linkage of H and L chains into complete tetrameric H₂L₂ antibodies. The co-expression can occur by using either the same or different plasmids in the same host. Genes for both H and L chains can be placed into the same plasmid, which is then transfected into cells, thereby selecting directly for cells that express both chains. Alternatively, cells may be transfected first with a plasmid encoding one chain, for example the L chain, followed by transfection of the resulting cell line with an H chain plasmid containing a second selectable marker. Cell lines producing H₂L₂ molecules via either route could be transfected with plasmids encoding additional copies of H, L, or H plus L chains in conjunction with additional selectable markers to generate cell lines with enhanced properties, such as higher production of assembled H₂L₂ antibody molecules or enhanced stability of the transfected cell lines.

In addition to mAbs or chimeric antibodies, the present invention is also directed to an anti-idiotypic (anti-Id) antibody specific for V region epitopes of the mAb antibody or

chimeric antibody of the invention. An anti-Id antibody is an antibody which recognizes unique determinants generally associated with the antigen-binding region of another antibody. The antibody specific for CCAA, such as 33.28, is termed the idiotype or Id antibody. The anti-Id can be prepared by immunizing an animal of the same species and genetic type mouse strain) as the source of the Id antibody with the Id antibody or the antigen-binding region thereof. The immunized animal will recognize and respond to the idiotype determinants of the immunizing antibody and produce an anti-Id antibody. The anti-Id antibody may also be used as an "immunogen" to induce an immune response in yet another animal, producing a so-called anti-anti-Id antibody. The anti-anti-Id may be epitopically identical to the original antibody which induced the anti-Id. Thus, by using antibodies to the idiotype determinants of a mAb, it is possible to identify other clones expressing antibodies of identical specificity.

Accordingly, the mAbs or chimeric antibodies of the present invention may be used to induce anti-Id antibodies in suitable animals, such as BALB/c mice. Spleen cells from such immunized mice can be used to produce anti-Id hybridomas secreting anti-Id mAbs. Further, the anti-Id mAbs can be coupled to a carrier such as keyhole limpet hemocyanin (KLH) and used to immunize additional BALB/c mice. Sera from these mice will contain anti-anti-Id antibodies that have the binding properties of the original mAb specific for a CCAA epitope.

The antibodies of the present invention, including their antigen-binding fragments and derivatives, have a multitude of uses relating to the diagnosis, monitoring and therapy of colon, breast, and ovarian cancer. Such uses are summarized in Schlom, J., *Canc. Res.*, 46:3225-3238 (1986), which is hereby incorporated by reference.

In diagnosis, the antibodies may be used in immunoassays (described below) to screen body fluids, such as serum, sputum, effusions, urine, cerebrospinal fluid, and the like, for the presence of CCAA. The antibodies may be used for scanning or radioimaging, when labelled with an appropriate radiolabel, to detect primary or metastatic foci of tumor cells. Furthermore, the antibodies are useful in lymphoscintigraphy to detect lymph node involvement in the disease.

The antibodies of the present invention are also useful for immunopathological analysis, such as the differential diagnosis of tumor type, the subclassification of the tumor based on its expression of CCAA. Such determinations would be important in assessment of metastatic potential, predicted responses to therapy and prognosis.

In particular, because of the specificity of the mAbs and chimeric antibodies of the present invention, they may permit the definition of defining subpopulations of tumor cells among the heterogeneous cells present in a growing tumor. These antibodies could therefore be used in the typing and cross-matching of the tumor cell "lines" comprising the tumor by means of flow cytometry, both at the time of surgery and prior to therapy. An analysis of the tumor cell subpopulations with the antibodies of this invention, and a battery of additional mAbs, is used to define (a) which antigen preparation would be the most appropriate for specific active immunotherapy, (b) which mAb or chimeric antibody would be efficacious for ADCC, and (c) which antibody or combination of mAbs should be used for imaging the patient at a later date in search for recurrent or metastatic tumors.

In addition to their diagnostic utility, the antibodies of the present invention are useful for monitoring the progression of disease by screening body fluids for CCAA, radioimaging

or suspensions, including saline and buffered media, parenteral vehicles including sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, such as those based on Ringer's dextrose, and the like. Preservatives and other additives may also be present, such as, for example, antimicrobials, antioxidants, chelating agents, and inert gases and the like. See, generally, *Remington's Pharmaceutical Science*, 16th ed., Mack Publishing Co., Easton, Pa., 1980.

In particular, the antibodies, fragments and derivatives of the present invention are useful for treating a subject having or developing colon, breast, and ovarian adenocarcinoma. Such treatment comprises parenterally administering a single or multiple doses of the antibody, fragment or derivative, or a conjugate thereof.

The antibodies of this invention can be adapted for therapeutic efficacy by virtue of their ability to mediate ADCC and/or CDC against cells having CCAA associated with their surface. For these activities, either an endogenous source or an exogenous source of effector cells (for ADCC) or complement components (for CDC) can be utilized.

The antibodies of this invention, their fragments, and derivatives can be used therapeutically as immunoconjugates (see for review: Dillman, R. O., *Ann. Int. Med.* 111:592-603 (1989)). They can be coupled to cytotoxic proteins, including, but not limited to, Ricin-A, Pseudomonas toxin, Diphtheria toxin, and tumor necrosis factor. Toxins conjugated to antibodies or other ligands are known in the art (see, for example, Olsnes, S. et al., *Immunol. Today* 10:291-295 (1989)). Plant and bacterial toxins typically kill cells by disrupting the protein synthetic machinery.

The antibodies of this invention can be conjugated to additional types of therapeutic moieties including, but not limited to, diagnostic radionuclides and cytotoxic agents such as cytotoxic radionuclides, drugs and proteins. Examples of radionuclides which can be coupled to antibodies and delivered in vivo to sites of antigen include ^{212}Bi , ^{131}I , ^{186}Re , and ^{90}Y , which list is not intended to be exhaustive. The radionuclides exert their cytotoxic effect by locally irradiating the cells, leading to various intracellular lesions, as is known in the art of radiotherapy.

Cytotoxic drugs which can be conjugated to antibodies and subsequently used for in vivo therapy include, but are not limited to, daunorubicin, doxorubicin, methotrexate, and Mitomycin C. Cytotoxic drugs interfere with critical cellular processes including DNA, RNA, and protein synthesis. For a fuller exposition of these classes of drugs which are known in the art, and their mechanisms of action, see Goodman, A. G., et al., *Goodman and Gilman's THE PHARMACOLOGICAL BASIS OF THERAPEUTICS*, 7th Ed., Macmillan Publishing Co., 1985.

The antibodies of this invention may be advantageously utilized in coordination with other monoclonal or chimeric antibodies, or with lymphokines or hemopoietic growth factors, etc., which serve to increase the number or activity of effector cells which interact with the antibodies.

The antibodies, fragments, or derivatives of this invention, attached to a solid support, can be used to remove soluble colon carcinoma-associated antigens from fluids or tissue or cell extracts. In a preferred embodiment, they are used to remove soluble tumor antigens from blood or blood plasma products. In another preferred embodiment, the antibodies are advantageously used in extracorporeal immunoadsorbent devices, which are known in the art (see, for example, *Seminars in Hematology*, Vol. 26 (2 Suppl. 1) (1989)). Patient blood or other body fluid is exposed to the

In addition to the pharmacologically active compounds, the new pharmaceutical compositions may contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Preferably, the preparations, contain from about 0.01 to 99 percent, preferably from about 20 to 75 percent of active compound(s), together with the excipient.

Preparations of the antibody, fragment or derivative of the present invention for parenteral administration, such as in detectably labelled form for imaging or in a free or conjugated form for therapy, include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propyleneglycol, polyethyleneglycol, vegetable oil such as olive oil, and injectable organic esters such as ethyloleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions

attached antibody, resulting in partial or complete removal of circulating CCAA (free or in immune complexes), of CCAA-bearing cells, following which the fluid is returned to the body. This immunoadsorption can be implemented in a continuous flow arrangement, with or without interposing a cell centrifugation step. See, for example, Terman, D. S. et al., *J. Immunol.* 117:1971-1975 (1976).

The present invention also provides the above antibodies, fragments and derivatives, detectably labelled, as described below.

The antibodies of the present invention are useful for immunoassays which detect or quantitate CCAA or cells bearing CCAA in a sample. Such an immunoassay typically comprises incubating a biological sample in the presence of a detectably labelled antibody of the present invention capable of identifying the tumor antigen, and detecting the labelled antibody which is bound in a sample.

Thus, in this aspect of the invention, a biological sample may be treated with nitrocellulose, or other solid support or carrier which is capable of immobilizing cells, cell particles or soluble proteins or glycoproteins. The support may then be washed with suitable buffers followed by treatment with the detectably labelled antibody of the present invention. The solid phase support may then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on said solid support may then be detected by conventional means.

By "solid phase support" or "carrier" is intended any support capable of binding antigen or antibodies. Well-known supports or carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, agaroses, and magnetite. The nature of the carrier can be either soluble to some extent or insoluble for the purposes of the present invention. The support material may have virtually any possible structural configuration so long as the coupled molecule is capable of binding to CCAA or the antibody specific for CCAA. Thus, the support configuration may be spherical, as in a bead, or cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface may be flat such as a sheet, test strip, etc. Preferred supports include polystyrene beads. Those skilled in the art will know many other suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

The binding activity of a given lot of antibody may be determined according to well-known methods. Those skilled in the art will be able to determine operative and optimal assay conditions for each determination by employing routine experimentation.

One of the ways in which the antibody of the present invention can be detectably labelled is by linking the same to an enzyme and use in an enzyme immunoassay (EIA) or enzyme-linked immunosorbent assay (ELISA). This enzyme, when subsequently exposed to its substrate, will react with the substrate generating a chemical moiety which can be detected, for example, by spectrophotometric, fluorometric or by visual means. In an alternate embodiment, the enzyme is used to label a binding partner for the antibody of the invention. Such a binding partner may be an antibody against the constant or variable region of the antibody of the invention, such as a heterologous anti-mouse immunoglobulin antibody. Alternatively, the binding partner may be a non-antibody protein capable of binding to the antibody of the present invention, such as staphylococcal protein A, or streptococcal protein G.

Enzymes which can be used to detectably label the CCAA-specific antibodies of the present invention, or the

binding partners for these antibodies, include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glycoamylase and acetylcholinesterase.

By radioactively labelling the antibody of the present invention or the binding partner, it is possible to detect CCAA through the use of a radioimmunoassay (RIA) (see, for example, Work, T. S. et al., *Laboratory Techniques and Biochemistry in Molecular Biology*, North Holland Publishing Company, N.Y. (1978)). The radioactive isotope can be detected by such means as the use of a gamma counter or a scintillation counter or by autoradiography. Isotopes which are particularly useful for the purpose of the present invention are well known in the art.

It is also possible to label the antibodies or binding partners with a fluorescent compound. When the fluorescently labelled antibody is exposed to light of the proper wave length, its presence can then be detected due to fluorescence. Among the most commonly used fluorescent labelling compounds are fluorescein isothiocyanate, rhodamine, phycocerythrin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine.

The antibodies can also be detectably labelled using fluorescence-emitting metals such as ^{152}Eu , or others of the lanthanide series. These metals can be attached to the antibody using such metal chelating groups as diethylenetriaminepentaacetic acid (DTPA) or ethylenediaminetetraacetic acid (EDTA).

The antibodies of the present invention also can be detectably labelled by coupling to a chemiluminescent compound. The presence of the chemiluminescently labelled antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction. Examples of particularly useful chemiluminescent labelling compounds are luminol, isoluminol, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound may be used to label the antibody, fragment or derivative of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems, in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labelling are luciferin, luciferase and aequorin.

Detection of the antibody, fragment or derivative may be accomplished by a scintillation counter, for example, if the detectable label is a radioactive gamma emitter, or by a fluorometer, for example, if the label is a fluorescent material. In the case of an enzyme label, the detection can be accomplished by colorimetric methods which employ a substrate for the enzyme. Detection may also be accomplished by visual comparison of the extent of enzymatic reaction of a substrate in comparison with similarly prepared standards.

In situ detection may be accomplished by removing a histological specimen from a patient, and providing the labelled antibody, or the unlabelled antibody plus a labelled binding partner to such a specimen. Through the use of such a procedure, it is possible to determine not only the presence of the antigen but also its distribution in the examined tissue. Using the present invention, those of ordinary skill will readily perceive that any of a wide variety of histological

methods (such as staining procedures) can be modified in order to achieve such in situ detection. Such methods include, for example, immunohistochemical staining procedures. In a preferred embodiment, an avidin-biotin immunoperoxidase staining system can be used, and a kit utilizing this system is also contemplated.

The kit employing mAbs or chimeric antibodies of the present invention can be used for immunohistochemical evaluation of colon, breast, and ovarian carcinoma. Indications for tissue study are to evaluate subpopulations of tumor cells that express the antigens defined by mAbs 31.1 and 33.28.

The colon kit is comprised of the reagents necessary for immunohistochemical analysis as follows:

- a) mAbs 31.1, 33.28 or mouse/human chimeric antibody Chi #1, and the mAb for carcinoembryonic antigen (CEA), the latter representing the standard monoclonal used for colon tissue immunohistochemistry;
- b) reagents for immunoperoxidase (blocking reagent) in the form of, for example, goat serum; and secondary antibody, such as, for example, goat anti-mouse antibody;
- c) immunoperoxidase; and
- d) reagents to produce the brown coloration.

Similar kits can be employed for the immunohistochemical analysis of breast and ovarian carcinoma.

The immunoperoxidase technique to be employed is that of Sternberger. The primary antibody (mAb or chimeric antibody) serves as an antigen which can bind more than one secondary antibody. The secondary antibodies form a "bridge" between the primary antibody and the horseradish peroxidase-antiperoxidase complexes.

The kit contemplated herein can be used to study fully developed colon, carcinoma, polyps in transformation to define the extent of malignant transformation, benign polyps to see if a site of transformation has been missed and inflammatory bowel disease to evaluate any sites of undetected transformation. Similar kits can be employed to study breast and ovarian carcinomas.

Another kit similar to the above kit is also contemplated which uses all five mAbs to colon carcinoma in order to evaluate all subpopulations of tumors and as such has the capability to type and cross-match the lesions.

The antibody, fragment or derivative of the present invention may be adapted for utilization in an immunometric assay, also known as a "two-site" or "sandwich" assay. In a typical immunometric assay, a quantity of unlabelled antibody (or fragment of antibody), is bound to a solid support that is insoluble in the fluid being tested and a quantity of detectably labelled soluble antibody is added to permit detection and/or quantitation of the ternary complex formed between solid-phase antibody, antigen, and labelled antibody.

Typical, and preferred, immunometric assays include "forward" assays in which the antibody bound to the solid phase is first contacted with the sample being tested to extract the tumor antigen from the sample by formation of a binary solid phase antibody-CCAA complex. After a suitable incubation period, the solid support is washed to remove the residue of the fluid sample, including unreacted tumor antigen, if any, and then contacted with the solution containing an unknown quantity of labelled antibody (which functions as a "reporter molecule"). After a second incubation period to permit the labelled antibody to complex with the CCAA bound to the solid support through the unlabelled antibody, the solid support is washed a second time to remove the unreacted labelled antibody. This type of forward

sandwich assay may be a simple "yes/no" assay to determine whether CCAA is present or may be made quantitative by comparing the measure of labelled antibody with that obtained for a standard sample containing known quantities of the antigen. Such "two-site" or "sandwich" assays are described by Wide (*Radioimmune Assay Method*, Kirkham, ed., E. & S. Livingstone, Edinburgh, 1970, pp. 199-206).

Other type of "sandwich" assays, which may also be useful with CCAA, are the so-called "simultaneous" and "reverse" assays. A simultaneous assay involves a single incubation step wherein the antibody bound to the solid support and labelled antibody are both added to the sample being tested at the same time. After the incubation is completed, the solid support is washed to remove the residue of fluid sample and uncomplexed labelled antibody. The presence of labelled antibody associated with the solid support is then determined as it would be in a conventional "forward" sandwich assay.

In the "reverse" assay, stepwise addition first of a solution of labelled antibody to the fluid sample, followed by the addition of unlabelled antibody bound to a solid support after a suitable incubation period, is utilized. After a second incubation, the solid phase is washed in conventional fashion to free it of the residue of the sample being tested and the solution of unreacted labelled antibody. The determination of labelled antibody associated with a solid support is then determined as in the "simultaneous" and "forward" assays. In one embodiment, a combination of antibodies of the present invention specific for separate epitopes may be used to construct a sensitive three-site immunoradiometric assay.

For purposes of in vivo imaging of colon, breast, and ovarian cancer using the antibodies of the present invention, there are many different labels and methods of labelling known to those of ordinary skill in the art. Examples of the types of labels which can be used in the present invention include radioactive isotopes, paramagnetic isotopes, and compounds which can be imaged by positron emission tomography (PET). Those of ordinary skill in the art will know of other suitable labels for binding to the antibodies used in the invention, or will be able to ascertain such, using routine experiments. Furthermore, the binding of these labels to the antibody can be done using standard techniques common to those of ordinary skill in the art.

For diagnostic in vivo imaging, the type of detection instrument available is a major factor in selecting a given radionuclide. The radionuclide chosen must have a type of decay which is detectable by a given type of instrument. In general, any conventional method for visualizing diagnostic imaging can be utilized in accordance with this invention.

Another important factor in selecting a diagnostic radionuclide for in vivo imaging is that the half-life of a radionuclide be long enough so that it is still detectable at the time of maximum uptake by the target issue, but short enough so that deleterious radiation of the host is minimized. In one preferred embodiment, a radionuclide used for in vivo imaging does not emit particles, but produces a large number of photons in a 140-200 keV range, which may be readily detected by conventional gamma cameras.

For in vivo diagnosis, radionuclides may be bound to the antibody either directly or indirectly by using an intermediary functional group. Intermediary functional groups which are often used to bind radioisotopes which exist as metallic ions to the antibodies are the chelating agents, diethylene triamine pentaacetic acid (DTFA) and ethylene diamine tetraacetic acid (EDTA). Examples of metallic ions which can be bound to the antibodies of the present inven-

tion are ^{99m}Tc , ^{123}I , ^{111}In , ^{131}I , ^{97}Ru , ^{67}Cu , ^{67}Ga , ^{125}I , ^{68}Ga , ^{72}As , ^{89}Zr , and ^{201}Tl .

The specifically exemplified mAbs 33.28 and 31.1, and the chimeric antibody Chi #1 may be used to facilitate the production of additional mAbs which bind the same or immunologically cross-reactive colon carcinoma-associated antigens. First, these antibodies may be conjugated to a chromatographic support, and used to immunopurify colon carcinoma-associated antigens. These purified antigens, in turn, may be used to stimulate an immune response in suitable animals. Secondly, spleen cells from the responsive animals may be fused to immortalizing cells, and the resulting hybridomas screened for secretion of antibodies which bind to the purified antigen and/or whose binding to colon carcinoma-associated antigen is competitively inhibited by antibody 33.28 or 31.1, or chimeric antibody Chi #1.

Having now generally described the invention, the same will be further understood by reference to certain specific examples which are included herein for purposes of illustration only and are not intended to be limiting unless otherwise specified.

EXAMPLE I

Preparation and Characterization of the Colon Carcinoma-Associated Antigen (CCAA)

The antigenic preparation was obtained from pooled colon carcinoma membranes according to the method described by Hollinshead et al., *Cancer* 56:480 (1985). This antigenic material was purified to the extent that the membrane fractions were free of HL-A antigens and were separated from much of the non-immunogenic glycoprotein fractions. In its final form the antigenic preparation was shown to be immunogenic in a specific manner in humans as evidenced by its capability of eliciting a delayed hypersensitivity reaction only in patients with active colon, breast, and ovary carcinoma.

Tumor cell suspensions in saline were prepared from fresh operating room specimens. Single cell suspensions, obtained by conventional means, were centrifuged for 10 minutes at about 400xg and the supernatant was retained. The cell pellet was resuspended and recentrifuged. The membrane material was examined by electron microscopy to assure that only membrane material (and no intact cells) was present, and the protein content was measured by the Lowry method.

The membrane material was next subjected to sequential low frequency sonication and resuspended as a soluble pool of membrane proteins. The soluble sonicates were separated by gel filtration on Sephadex-6200. Fractions of 2 ml were collected and the absorbance profile at 220 and 280 nm was recorded. Fractions comprising individual protein peaks were pooled, and the pools were concentrated by Diaflo ultrafiltration. Sephadex-G200 fractions IB and IIA, as defined by Hollinshead et al., (supra), were further purified by gradient polyacrylamide gel electrophoresis (PAGE). The fractions were tested for their ability to elicit positive delayed cutaneous hypersensitivity reactions in patients with colon carcinoma. Those fractions with immunogenic activity were said to contain colon carcinoma-associated antigens (CCAA) and were employed as immunogens and screening agents in the preparation of the mAbs.

By gradient PAGE, a double-banded antigen distinct from that of carcinoembryonic antigen (Gold, P. et al., *J. Exp. Med.* 122:467-481 (1965); Hollinshead, A. et al., *Cancer* 56:480 (1985)) was identified and isolated. The bands comprising this antigen migrated 6.3 and 6.6 cm. distant from

tracking dye. Biochemical analysis of the antigen proved that it was glycoprotein. The molecular weight of the antigen was estimated based on the electrophoretic mobility of transferrin (6.4-6.5 cm) which has a molecular weight of 76.5 kDa.

EXAMPLE II

Preparation and Screening of Monoclonal Antibodies

Monoclonal antibodies (mAbs) against human colon carcinoma-associated antigens (CCAA) were obtained by the production and cloning of hybrids resulting from the fusion of mouse myeloma cells Sp2/0-Ag14 with spleen cells from BALB/c mice which had been immunized with the CCAA described above.

Five hybrid clones were established, as described below, and designated as 31.2, 31.1, 77, 33.23 and 33.28. All five mAbs reacted strongly with the CCAA and with two colon carcinoma cell lines (SW480 and SW620) when assayed by ELISA. Two of the mAbs, 31.1, and 33.28, were studied in greatest detail.

A. Immunization and Cell Fusion

BALB/c mice were immunized by intraperitoneal injection of 50 μg of the CCAA described above emulsified in complete Freund's adjuvant, as described by Hollinshead in clinical trials (Hollinshead et al., supra). Ten days later the mice received an intravenous booster injection of the same amount of CCAA in saline. Mice were sacrificed three days later and their spleen cells obtained. Cell fusion was performed by incubation 5×10^7 mouse spleen cells with 10^7 sP2/0-Ag14 myeloma cells in 40% polyethylene glycol (MW=1500).

B. Screening of Hybrid Clones

An enzyme-linked immunosorbent assay (ELISA), described by Tsang et al., *JNCI* 77:1175 (1986), was used for the detection of hybridoma clones producing antibodies specific for the CCAA. CCAA (100 n/well) was immobilized on polystyrene microplates. Antibodies present in the test supernatants were allowed to bind to the immobilized antigens. The presence of the bound murine mAbs was detected with peroxidase-conjugated second antibodies, specific for mouse immunoglobulins, followed by the chromogenic substrate for peroxidase, O-phenyldiamine. Wells showing color reactions yielding Absorbances ≥ 0.500 units were scored as positive. Negative controls gave values of 0.01 to 0.09 units.

Hybridoma wells scoring as positive by ELISA were further screened by indirect immunofluorescence, using various tumor cells and normal cells as identified in Table 1, below. All of the tumor cell lines were obtained from the ATCC. Cells were incubated with hybridoma culture supernatants at an appropriate dilution (1:2) in phosphate buffered saline (PBS) for 1 hour at 4° C. The cells were washed and incubated with a fluorescein-labelled goat anti-mouse immunoglobulin antibody. The cells were then washed three times with PBS and examined by fluorescence microscopy. The results appear in Table 1.

TABLE 1

INDIRECT IMMUNOFLUORESCENCE REACTIVITY OF ANTI-COLON CARCINOMA-ASSOCIATED ANTIGEN(S) (CCAA) MoAbs WITH HUMAN CULTURE CELLS ^a					
CELLS	REACTIVITY OF MoAbs ^b				
	31.2	77	31.1	33.28	33.23
TUMOR LINES					
SW948 (COL)	-	+	-	-	-
HCT116 (COL)	-	-	-	+	-
WIDR (COL)	+	+	+	+	+
COLO320 (COL)	+	+	+	-	-
HS619 (COL)	-	-	-	-	-
HS853 (COL)	-	-	-	-	-
CACO-2 (COL)	+	+	-	-	-
SK-CO-1 (COL)	+	+	-	+	+
HT-29 (COL)	+	+	-	+	+
SW1116 (COL)	+	-	-	+	+
SW480 (COL)	+	+	+	+	+
SW620 (COL)	+	+	+	+	+
Z31 (BR)	-	-	-	-	-
CAMA-1 (BR)	-	-	-	-	-
PAN-1 (PAN)	-	-	+	-	-
MIA (PAN)	-	-	+	-	-
HS766T (PAN)	-	-	-	-	-
M-14 (MEL)	-	-	-	-	-
HT1080 (FIB)	-	-	-	-	-
LM (OS)	-	-	-	-	-
TE-85 (OS)	-	-	-	-	-
NORMAL SKIN	-	-	-	-	-
FIBROBLAST	-	-	-	-	-
BONE MARROW	-	-	-	-	-
CELL	-	-	-	-	-
NORMAL HUMAN	-	-	-	-	-
PBMC	-	-	-	-	-

^aCULTURE SUPERNATANT WAS DILUTED 1:2 WITH PBS.

^bPOSITIVE (+) AND NEGATIVE (-) REACTIVITIES WERE DEFINED BY INTENSITY OF THE MEMBRANE FLUORESCENCE AS COMPARED TO THE BACKGROUND.
C. COL: COLON CARCINOMA; BR: BREAST CARCINOMA; PAN: PANCREATIC CARCINOMA; FIB: FIBROSARCOMA.

EXAMPLE III

Analysis of Monoclonal Antibodies and their Reactivity

The anti-CCAA mAbs produced and detected as above were also tested for reactivity with fresh human tissue. Cryostat sections of the tissue types listed in Table 2, below, were fixed with 3.5% formaldehyde in PBS and then washed three times with PBS. For indirect immunofluorescence studies, the sections were incubated with the mAbs and then stained with a fluorescein-labelled second antibody as above.

As is shown in Table 2, mAbs 31.1 and 33.28 were highly specific for colon carcinoma cells. This indicates that an antigen (CCAA) which was highly specific for colon carcinoma and, furthermore, was immunogenic in colon carcinoma patients (positive DH reactions), and served as a successful immunogen in mice for the devnd phycoerythrin excitation was used. Trigger regions were established by examining cells by forward versus 90° light scatter. As shown in Table 4, both 31.1 and 33.28 bound to colon carcinoma cells; neither mAb bound significantly to PBMC.

TABLE 2

Indirect Immunofluorescence of Anti-CCAA Mabs with Fresh Human Tissues ^a		
Tissues	Reactivity of Mabs	
	33.28	31.1
Tumor		
Colon Carcinoma	3/3	3/3
Pancreatic Carcinoma	0/2	0/2
Melanoma	0/2	0/1
Breast Carcinoma	0/2	0/1
Normal		
Placenta	0/1	0/1
Liver	0/1	0/1
Colon	0/3	0/3
Spleen	0/1	0/1
Thymus	0/1	0/1
Muscle	0/1	0/1

^aAscitic fluid was diluted 1:50 with PBS. Cryostat sections (4-6 μm thick) were fixed with 3.5% formaldehyde in PBS for 10 minutes and then washed three times in PBS. Sections were stored at -70° C. unless used immediately. Results are expressed as number of positive/negative of tissue tested.

Table 3 shows the results of an immunoabsorption analysis of the Mabs. Three colon carcinoma cell lines (HT-29, WIDR and SW 620) and an osteosarcoma cell line (LM) were used to absorb fluorescein isothiocyanate (FITC)-conjugated Mabs 31.1 or 3.28. Ascites fluid from mice in which the hybridomas were growing, diluted 1:50, was added to the absorbing cells. The mixtures were incubated for 1 hr at 4° C. Either 2×10⁷ cells (Table 3, Part A) or 10⁴ cells (Table 3, Part 5) were used to absorb the antibodies (Table 3) The osteosarcoma cell line did not absorb out 31.1 or 33.28 activity, while the colon carcinoma cell lines did.

TABLE 3

Immunoabsorption Analysis of mAbs								
mAb	Absorbing Cells							
	HT-29		WIDR		SW620		LM	
	A	B	A	B	A	B	A	B
33.28	-	+	-	+	-	+	+	+
31.1	-	+	-	+	-	+	+	+

Cytofluorometric analysis was used to measure the binding of 31.1, 33.28 and a control mAb to HT29, WIDR and SW480 tumor cells and to peripheral blood mononuclear cells (PBMCs) An Ortho Spectrum III Cytofluorograph, equipped with an argon laser capable of fluorescein and phycoerythrin excitation was used. Trigger regions were established by examining cells by forward versus 90° light scatter. As shown in Table 4, both 31.1 and 33.28 bound to colon carcinoma cells; neither mAb bound significantly to PBMC.

TABLE 4

SUMMARY OF CYTOFLUOROMETRIC ANALYSIS			
% of cells stained with mAb:			
Cells	33.28	31.1	Control
HT29	51.0	53.9	9.2
WiDr	21.0	ND	8.1
SW480	37.0	32.0	3.8
PBMC	2.1	2.1	ND

The heavy and light chain isotypes of the mAbs were determined by immunodiffusion. The 31.1 mAb was found to be an IgG1 with a kappa light chain. The 33.28 mAb was found to be an IgG2a with a kappa light chain (Table 5). This is in strong contrast to the prior art mAb 19.9 (Herlyn, M. et al., *J. Biol. Chem.* 257:14365-14369 (1982)) which is of the IgG1 class (Herlyn, D. et al., *Proc. Natl. Acad. Sci. USA* 79:4761-4765 (1982)). Importantly, antibodies of the IgG2a class are expected to be more useful for immunotherapeutic purposes (Colcher, D. et al., *Proc. Natl. Acad. Sci. USA* 78:3199-3203 (1981)). Although the 19.9 mAb has reactivity to colon tumors, it was derived by immunization with pancreatic carcinoma antigens, and is therefore cross-reactive with colon. This is analogous to the situation with the B72.3 mAb ([citation]), which is a colon-reactive antibody obtained by immunization with breast cancer tissue.

TABLE 5

Culture	Isotyping of Monoclonal Antibodies					
	Light Chains					
Supernatant	IgG1	IgG2a	IgG2b	IgM	Kappa	Lambda
31.2	-	+	-	-	+	-
31.1	+	-	-	-	+	-
77	-	-	+	-	+	-
33.23	-	+	-	-	+	-
33.28	-	+	-	-	+	-

EXAMPLE IV

Characterization of the Colon Carcinoma-Associated Antigen

The molecular mass of the antigens to which the above mAbs bound was determined by Western blot analysis using soluble protein extracted from colon carcinoma cell lines SW480 and SW620. The 33.28 and 31.1 mAbs reacted with molecules having apparent molecular weights of 61.1 kDa and 72 kDa, respectively, from both of these cell lines. These mAbs did not react with material from human PBMCs or from human tumor cell lines of other histologic types in Western blot analysis.

In order to define better the specificity of the mAbs of the present invention for the immunizing CCAA and to establish whether the mAbs reacted with an immunogenic component of the cell membrane preparation which has been used in clinical immunotherapy trials (Hollinshead et al., *supra*), the original immunogenic preparation described above was performed by high performance liquid chromatography (HPLC).

The analysis revealed 4 distinct peaks, each of which was tested for immune reactivity (elicitation of DH) in patients with colon carcinoma by skin test (FIG. 1). Among the 10

patients with colon carcinoma tested only the material in peak #4 induced a cutaneous DH reaction. The peak #4 antigen was found to react with mAb 33.28, while mAb 31.1 reacted with peak #3, the next most prominent peak.

The references cited above are all incorporated by reference herein.

EXAMPLE V

Affinity Purification of Colon Carcinoma-Associated Antigen

The mAb 33.28 was used in affinity chromatography to isolate antigen extracted from cells of the HT-29 line. Five mg of purified 33.28 IgG was coupled to CNBr-activated sepharose 4B. The column was pre-eluted with 0.05M diethylamine, pH 11.5, and then equilibrated with 0.14M NaCl/0.01M Tris (pH 8.0). CCAA preparation was applied to the column, and the column was eluted with 0.05M diethylamine, pH 11.5. The eluted fractions were neutralized by the addition of 1M Tris-HCl, pH 8.0.

The material bound and eluted from the 33.28 affinity matrix was then subjected to HPLC. The eluted CCAA preparation was adjusted in starting buffer (0.01M sodium phosphate buffer, pH 7.0), applied to a Synchropak Wax weak anion exchange HPLC column (250x4.6 mm) and eluted with a gradient of 0 to 1M NaCl in 0.01M sodium phosphate buffer, pH 6.0, at a flow rate of 1 ml/min. Anion exchange chromatography was performed using a Hewlett-Packard HPLC (HP 1090, Hewlett-Packard, Arondale, Pa.).

Results appear in FIG. 2. The antigenic material derived from HT-29 cells isolated by mAb 33.28 gave a peak that matched peak #4 described above and had similar immunogenic activity in humans, indicating the utility of mAb 33.28 for isolation of a colon cancer preparation which is immunogenic for humans.

EXAMPLE VI

ADCC Activity of mAbs 33.28 and 31.1

In order to be therapeutically useful, a mAb specific for an immunogenic tumor antigen should have the following properties: (a) high tumor tissue specificity, (b) absence of cross-reactivity to normal human tissue, and (c) a biological activity associated with destruction of tumors, such as antibody-dependent cellular cytotoxicity (ADCC).

The ADCC activity of mAbs 33.28 and 31.1 was tested on the colon carcinoma line WiDr as target cell. The melanoma cell line, M-14, served as a specificity control. ADCC was assayed using a conventional 4 hr. ⁵¹Cr release assay using normal human PBMC as effector cells, and the results are shown as percent isotope release (% lysis) (Table 6). The background lysis was 8.3%. At an effector:target ratio of 100:1, mAb 33.28 caused 40.3% lysis of tumor cells, and 31.1 induced 51.8% lysis.

TABLE 6

ADCC Activity of mAbs 33.28 and 31.1						
Antibody	% Lysis of Target Cells at E:T Ratios:					
	WDR			M-14		
or Control	25	50	100	25	50	100
33.28	23.1	40.3	45.3	6.9	8.4	9.0
31.1	14.3	26.7	51.8	7.5	6.4	8.7
OSA1	10.0	9.2	12.2	11.4	14.8	10.9
NMS	12.2	11.7	13.1	14.2	15.0	11.1
PBS	8.2	5.1	7.6	11.0	14.2	10.5

ADCC was assayed by a 4 hour ^{51}Cr release assay. Background ^{51}Cr release was 8.3%. E:T Ratio indicates effector cell-to-target cell ratios. The mAb or serum was tested at a 1:100 dilution; OSA1 - mAb to osteosarcoma associated antigens; NMS - normal mouse serum; WDR - colon carcinoma cell line; M14 - melanoma cell line.

EXAMPLE VII

Detection of Circulating CCAA with mAbs 33.28 and 31.1

The mAbs of the present invention were tested for their ability to detect circulating CCAA in 79 unknown serum samples (Table 7). The assay was based on the ability of the serum samples to inhibit binding of the mAb to the CCAA in ELISA. None of the 50 normal serum samples gave false positive results. Nine of the ten serum samples from patients with active colon carcinoma were positive. None of the sera from disease-free colon cancer patients one year post-resection were positive.

TABLE 7

Detection of Circulating Colon Carcinoma-Associated Antigen					
DONOR CONDITION	No. of Samples	No. of sera inhibiting binding of mAbs:			
		33.28		31.1	
		<15% (Neg)	>15% (Pos)	<15% (Neg)	>15% (Pos)
Colon Carcinoma	10	3	7	2	8
Colon Carcinoma (Resected)	4	4	0	4	0
Breast Carcinoma	9	9	0	9	0
Melanoma	5	5	0	5	0
Prostate Cancer	1	1	0	1	0
Normal Serum	50	50	0	50	0

Colon carcinoma-associated antigens was detected by ELISA. 100 μl of serum were used in each assay.

EXAMPLE VIII

Comparison of Specificity with Other mAbs Reactive with Colon Tumors

Further studies of tumor specificity were conducted using ELISA (Table 8). The mAb 31.1 was compared with CC49, a colorectal carcinoma-specific mAb purified from B-72.3, and a control mouse myeloma protein. 31.1 was shown to react with a narrower range of colorectal carcinomas than did CC49. However, it had a higher degree of specificity, having lower or no cross-reactivity with stomach tumors or normal colon tissue.

TABLE 8

ELISA ON NORMAL AND TUMOR TISSUES USING MAb 31.1, CC49 AND MOPC-21

Tissues	31.1	CC49	MOPC-21
Colorectal Carcinomas			
1. COCA2A	-	+++	-
2. COCA2	-	+++	-
3. COCA3	+	++	-
4. COCA4	+++	+++	-
5. G820	±	++	-
6. G853	+++	++	-
7. G817	+++	+++	-
8. G781	-	-	-
Other Carcinomas			
1. Breast CA1	-	-	-
2. Breast CA2	-	-	-
3. Lung CA1	-	-	-
4. Lung CA2	-	-	-
5. Ovarian CAD106	-	-	-
6. Ov CA5	-	-	-
7. Ov CAV5	-	-	-
8. Ov CAV45	-	-	-
9. Ov CAV43	-	-	-
10. Stomach CA14A	++	+++	-
11. Stomach CA12A	-	+++	-
12. Stomach CA15A	-	-	-
Other Normal Tissues			
1. Endometrium E21	-	-	-
2. Endometrium EC19'	-	-	-
3. Endometrium EC17	-	+	-
4. Endometrium EC18 (RBC)	-	-	-
5. Red blood cells 1	-	-	-
6. RBC 2	-	-	-
7. RBC 3	-	-	-
8. RBC 4	-	-	-
9. RBC 5	-	-	-
10. RBC 6	-	-	-
11. RBC 7	-	-	-
12. RBC 8	-	-	-
13. RBC 9	-	-	-
14. RBC 10	-	-	-
15. RBC 11	-	-	-
16. Granulocytes	-	-	-
17. 385	-	-	-
18. 386	-	-	-
19. Normal spleen 3	-	-	-
20. 392 (N. Spleen)	-	-	-
21. 395 (N. Liver)	-	-	-
22. 387 (N. Kidney)	-	-	-
23. 398 (N. Spleen)	-	-	-
24. 390 (N. Liver)	-	-	-
25. N. Spleen #1	-	-	-
26. N. Spleen #2	-	-	-
27. 800 (N. Colon)	-	++	-
28. N. Colon (GW)	-	++	-
29. N. Colon (Meloy)	-	-	-
30. N. Colon	-	-	-
31. G1155B (N. Colon)	-	-	-
32. G1164B (N. Colon)	-	-	-
33. N. Colon	±	-	-
34. Normal Stomach A	-	-	-
35. N. Stomach B	-	-	-
36. N. Stomach C	-	-	-
37. Normal Lung	-	-	-
38. Normal Liver	-	-	-

CC49 - NCI monoclonal antibody to colorectal carcinomas

MOPC-21 - negative control myeloma protein

All monoclonal antibodies were used at 40 ng/well, POGAM at 1:3000 dilution

EXAMPLE IX

In Vivo Localization of mAbs 33.28 and 31.1 to Tumors

The in vivo behavior of the mAbs of the present invention was examined by pharmacokinetic studies using ¹²⁵I-labelled mAb and athymic nude mice bearing LS-174T colon tumor xenografts. Mice implanted with the A375 melanoma were employed as controls. The relative concentration of mAb in tumor as compared to adjacent normal tissue such as liver and spleen is represented by the radiolocalization index (concentration of radiolabelled material in tumor/concentration in surrounding tissue). The biodistribution of ¹²⁵I-labelled 31.1 and 33.28 mAbs is shown in Table 9. Both mAbs were able to significantly concentrate within the tumor, compared to localization in normal tissues (spleen and liver). This selective accumulation was six-fold at 96 hours and about 12-fold at 168 hrs. The radiolocalization indices for both mAbs to tumor as compared to blood, liver and spleen are shown in FIG. 3 and FIG. 4.

TABLE 9

Biodistribution of ¹²⁵ I-mAbs in Tumor-Bearing Athymic Nude Mice				
Tissue	96 hours		168 hours	
	LS174T	A375	LS174T	A375
A. mAb 31.1				
Blood	7.30	NA	4.67	4.42
Tumor	21.92	NA	25.43	2.91
Liver	3.74	NA	2.16	1.24
Spleen	3.68	NA	2.41	1.32
B. mAb 33.28				
Blood	7.81	NA	5.58	3.95
Tumor	13.12	NA	15.50	2.24
Liver	2.55	NA	1.74	1.68
Spleen	2.31	NA	1.70	1.92

Results are expressed in % injected dose/gram of tissue.
LS174T = colon carcinoma; A375 = melanoma.

EXAMPLE X

Immunohistochemical Studies

The ability to detect tumor markers in the serum, image the related neoplastic process and define the cell population of that neoplasm by immunohistochemistry depends on the ability of specific mAbs to selectively characterize a tumor population.

The mAbs 31.1 and 33.28 were tested in more than 50 colon carcinomas by means of immunoperoxidase staining. They have been found to be highly reactive with the colon neoplasm and did not interact with the adjacent normal tissues. When polyps were evaluated, the wholly benign lesions such as villo-tubular adenomas showed no reactivity. Villous adenomas undergoing transformation reacted only at the site of malignancy. When similar tissues were evaluated using the more common carbohydrate-antigen derived mAbs, normal adjacent colon tissue reacted equally with the neoplastic portion of the specimen. With the 31.1 and 33.28 mAbs, each appeared to stain different cell populations within the tumor. This suggests that surface antigens representing different oncogene products were being defined.

EXAMPLE XI

Selective Binding to Subpopulations of Epithelial Cells in Paraffin-Embedded Benign and Malignant Mammary Lesions

Forty-one formalin-fixed, paraffin-embedded benign and malignant breast specimens were studied with mAbs 31.1

and 33.28, using the avidin-biotin staining method. Without enzymatic pretreatment, positive staining of epithelium was observed on the cell surface and in the cytoplasm with both antibodies. 7/21 (33%) duct carcinogens were positive with mAb 31.1 as were 5/20 (25%) samples of benign breast disease. 10/21 (48%) duct carcinomas were positive with mAb 33.28 together with 7/20 (35%) specimens of benign mammary disease. 10 to 75% of the cell population was positively stained. These results indicate that the antigens defined by mAbs 31.1 and 33.28 are expressed in a select group of women with breast disease and would be useful for diagnosis of said disease.

EXAMPLE XII

Selective Binding to Subpopulations of Epithelial Cells in Fresh Frozen Benign and Malignant Ovarian Tumors

Fresh frozen tissue biopsies obtained from twenty-one ovarian tumors subjected to immunocytochemical analysis were studied with mAbs 31.1 and 33.28 using the avidin biotin indirect immunoperoxidase assay. Focal positive staining was observed in 4/7 papillary mucous, 1/1 mucinous and 1/2 endometrioid adenocarcinoma. None of the nonepithelial ovarian tumors stained positive using these monoclonal antibodies. These results indicate that the antigens as defined by mAbs 31.1 and 33.28 are expressed in a select group of woman with ovarian cancer and would be useful for diagnosis of said disease.

EXAMPLE XIII

Immunoreactivity of Human Carcinoma Tissues and Cell Lines with mAbs 31.1 and 33.28

The mAbs 31.1 and 33.28 were used to screen a panel of cell lines including colon adenocarcinoma, lymphoma, leukemia and neuroblastoma lines.

Using the avidin-biotin immunoperoxidase staining system, the mAbs 31.1 and 33.28 were shown to strongly bind to colon adenocarcinoma cell lines WIDR and HT-29. Immunoreactions were not observed with KG1-a, HL-60, Molt-3 and JUKRAT cell lines. Both antibodies reacted weakly with one lymphoma line (JY). The mAb 33.28 reacted weakly with one leukemic line (K562) and a neuroblastoma line (U87.MG). These results confirm and extend previous flow cytometry and immunofluorescent results in which it has been reported that strong binding reactions were observed with these mAbs with colon adenocarcinoma cell lines and reactions were not observed with other tumor cell lines. Using flow cytometry, mAb 31.1 reacted with 85% of HT-29 and WIDR colon carcinoma cells but not with SKBR-3 breast cancer cells.

Both antibodies were extensively shown to bind distinctively to colon carcinoma tissues (mAb 33.28-84%, mAb 31.1-64%), and not to normal tissues or malignant tissues including neuroblastoma tissues (0/3), lymphomas (0/3) and leukemic infiltrates (0/3) tested. These results suggest that these antibodies can serve as a useful research tool in evaluating tumor markers in cancer and cell biology research.

EXAMPLE XIV

Expression and Characterization of Chimeric Antibodies Against Human Colorectal Carcinoma-Associated Antigen

A chimeric mouse/human heavy chain gene was constructed by splicing the exon of the 31.1 antibody heavy